


REVIEW

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Biosynthesis of nanoparticles and silver nanoparticles

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Abstract

In this century, the development of nanotechnology is projected to be the establishment of a technological evolutionary of this modern era. Recently, nanotechnology is one of the most active subjects of substantial research in modern material sciences and hence metal nanoparticles have a great scientific interest because of their unique optoelectronic and physicochemical properties with applications in diverse areas such as electronics, catalysis, drug delivery, or sensing. Nanotechnology provides an understanding on fundamental properties of objects at the atomic, molecular, and supramolecular levels. Besides, nanotechnology also leads an alternative technological pathway for the exploration and revolution of biological entities, whereas biology provides role models and biosynthetic constituents to nanotechnology. The findings of this review are important to provide an alternative for the green synthesis of silver nanoparticles. It showed more cost-effective and environmental friendly application as well as easier for large production, with relation to the properties of silver nanoparticles as antimicrobial, can be served well as an alternative antiseptic agent in various fields. Typically, silver nanoparticles are smaller than 100 nm and consist of about 20–15,000 silver atoms. Due to the attractive physical and chemical properties of silver at the nanoscale, the development of silver nanoparticles is expanding in recent years and is nowadays significant for consumer and medical products.

Keywords: Bio-nanotechnology, Silver nanoparticles, Plant-mediated synthesis, Antimicrobial agent, Biosynthesis, Nanotechnology

Background

In recent years, nanotechnology is an escalating field of modern research (Edhaya Naveena and Prakash 2013) involving in synthesis design, characterization, production, and application of structures, devices, and systems by controlling shape and size at the nanometer scale (Madhuri et al. 2012). Nanotechnology also involves synthesis of nanoparticles of size ranging from 1 to 100 nm (EU 2011; Adlakha-Hutcheon et al. 2009). Moreover, there is a new branch of nanotechnology existing, which is bio-nanotechnology that integrates principles of biology with physical and chemical procedures to generate nano-sized particles with specific functions (Kathiresan

et al. 2009; Qi and Wang 2004; Roduner 2006). The bio-based protocols for synthesis of nano-metals are both environmentally and economically green as they are based on green chemistry principles and are simple, relatively inexpensive, and easily scaled up for larger scale production (Mohanpuria et al. 2008; Iravani 2011; Prabhu and Poulouse 2012). However, the chemical methods available are often expensive, utilize lethal chemicals, and are comparatively complex. Hence, biosynthesis of nanoparticles using biological agents such as microbes or plant extracts has gained much attention in the area of nanotechnology in last few decades (Malik et al. 2014). Generally, there are three main steps involved in green synthesis method, i.e., reaction medium selection, biological reducing agent selection, and selection of non-carcinogenic substances for stability of nanoparticles (El-Shishtawy et al. 2011). Yet, plant-mediated preparation of nanoparticles can be advantageous over other bio-based synthesis because the procedure of maintaining

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cell cultures can be omitted and it is also suitable for large scale production under non-aseptic environments (Makarov et al. 2014).

Silver nanoparticles are well-known antimicrobial agents in surgically implanted catheters in order to reduce the infections caused during surgery (Oei et al. 2012) and are proposed to possess anti-fungal, anti-inflammatory, anti-angiogenic, and anti-permeability activities. Silver is also one of the main components in the various creams for healing wounds (Saikia et al. 2015). However, silver nanoparticles are now being introduced as an alternative antibacterial agent replacing silver ions. Both silver ions and silver nanoparticles have inhibitory and lethal effects on bacterial species such as *Escherichia coli*, *Staphylococcus aureus*, and even yeast. But, the formation of complexes for silver ions is limited and the effect of the silver ions somehow remains only for a short period (Méndez-Vilas 2011). Yet, this drawback has been resolved by the application of the intact silver nanoparticles which have greater antibacterial properties by promoting the synthesis of reactive oxygen species such as hydrogen peroxide (Mohammed 2015). In addition to antibacterial activity of the silver nanoparticles, a complete disruption of the bacterial membrane of *Escherichia coli* cells was observed after few minutes in contact with silver nanoparticles under TEM analysis (Raffin et al. 2008). The high efficiency of silver nanoparticles is mainly due to the availability of larger surface area to volume ratio for interactions, easing the penetration and disruption of nanoparticles into the bacterial cells, as compared to micro-sized silver ions (Durán et al. 2010).

Preparation of Nanoparticles

Generally, nanomaterials can be fabricated through two main methods, i.e., “top-down” and “bottom-up” (Fig. 1)

approaches (Forough and Farhadi 2010). The top-down approach basically works with the material in its bulk form, and the size reduction to the nanoscale is then achieved by specialized ablations, e.g., lithography, thermal decomposition, laser ablation, mechanical milling, etching, and sputtering (Abou El-Nour et al. 2010). Alternatively, the “bottom-up” approach is more preferable for the preparation of nanoparticles, where involving a homogeneous system wherein catalysts (e.g., reducing agent and enzymes) synthesize nanostructures that are controlled by catalyst properties, reaction media, and conditions (e.g., solvents, stabilizers, and temperature). For instance, chemical reduction method is the most common synthetic pathway for metal nanoparticles synthesis (Pal et al. 2011). In the case of silver nanoparticles, the chemical reduction method is carried out based on the reduction of aqueous silver nitrate in an appropriate operating medium using chemical reductants such as sodium citrate or branched polyethylenimine. In this way, negatively charged silver nanoparticles can be obtained from the process using sodium citrate acting as reductant, while positively charged silver nanoparticles can be synthesized from the reaction with branched polyethylenimine as reductant (Moghaddam 2010). Thus, the physicochemical properties, surface, and morphological characteristics of nanoparticles can possibly be controlled depending on the subsequent application through variation in precursor concentrations and reaction conditions (Mason et al. 2012).

Green synthesis of nanoparticles

There are a variety of chemical and physical preparation methods available for the fabrication of nanoparticles including radiation, chemical precipitation, photochemical methods, electrochemical, and Langmuir–Blodgett

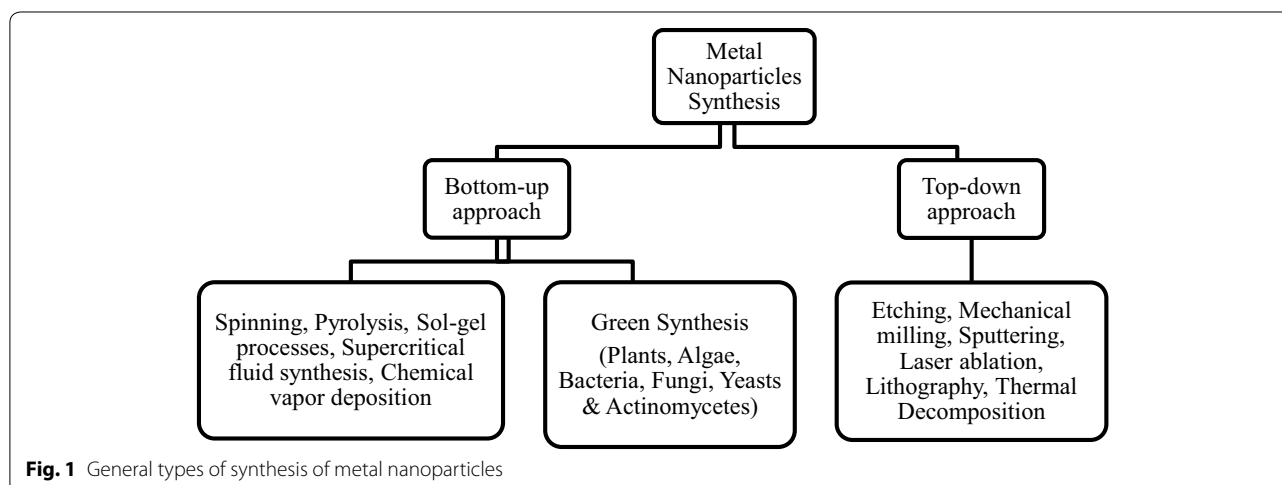


Table 1 Uses of various biological entities in the synthesis of silver nanoparticles during the period of 2009–2015

Nanoparticles produced	Biological entity	Size (nm)	References
AgNP	<i>Cycas circinalis</i> , <i>Ficus amplissima</i> , <i>Commelina benghalensis</i> , and <i>Lippia nodiflora</i>	13–51	Johnson and Prabu (2015)
AgNP	<i>Sinapis arvensis</i> seed exudates	1–35	Khatami et al. (2015)
AgNP	<i>Butea monosperma</i> leaf extract	5–30	Chaturvedi and Verma (2015)
AgNP	<i>Musa balbisiana</i> , <i>Azadirachta indica</i> and <i>Ocimum tenuiflorum</i>	20–50	Banerjee et al. (2014)
AgNP	<i>Quercus infectoria</i> extract	40	Heydari and Rashidipour (2015)
AgNP	<i>Schizophyllum commune</i> (mushroom fungus)	51–93	Arun et al. (2014)
AgNP	<i>Lantana camara</i> berry	75.2	Kumar et al. (2015)
AgNP	Pomegranate peel extract	5–50	Shanmugavadivu et al. (2014)
AgNP	<i>Aloe vera</i> extract	500	(SarahIbrahim et al. (2014)
AgNP	<i>Aegle marmelos</i> (Bael) fruit extract	34.7	Nithya Deva Krupa and Raghavan (2014)
AgNP	<i>Bacillus stearothermophilus</i>	14 ± 4	El-Batal et al. (2013)
AgNP	<i>Sargassum muticum</i> (brown marine macroalgae)	5–15	Azizi et al. (2013)
AgNP	<i>Streptomyces</i> sp. LK3	5	Karthik et al. (2014)
AgNP	<i>Glycine max</i> (soybean)	7–29	Sasikala et al. (2012)
AgNP	<i>Schizophyllum radiatum</i> HE863742.1	10–40	Metuku et al. (2014)
AgNP	<i>Tinospora cordifolia</i> leaf extract	3.3–20.6	Rajathi et al. (2012)
AgNP	<i>Fusarium oxysporum</i>	25–50	Birla et al. (2013)
AgNP	<i>Trichoderma reesei</i> (fungus)	5–50	Khabat Vahabi et al. (2011)
AgNP	<i>Sargassum longifolium</i>	30	Saraniya Devi et al. (2013)
AgNP	<i>Gliricidia Sepium</i>	10–50	Raut Rajesh et al. (2009)

techniques, but these methods are often extremely expensive and non-environmental friendly due to the use of toxic, combustible, and hazardous chemicals, which may pose potential environmental and biological risk and high energy requirement (Awwad et al. 2013). The drawbacks of low production rate, structural particle deformation, and inhibition of particle growth are also encountered in these nanoparticles synthesis. Currently, there is a growing need to develop sustainable preparation of nanoparticles that get rid of using harmful organic chemical substances, since noble nanoparticles are widely applied to areas of human contact (Shams et al. 2013). To achieve the principle of green chemistry process, it leads to in search of green synthesis of nanoparticles which can be done by five methods as below:

Polysaccharide method

By using polysaccharide method, metal nanoparticles are synthesized by using water and polysaccharides acting as a reducing agent, a stabilizing agent, or both reducing and stabilizing agents. For example, the fabrication of silver nanoparticles can be performed by using starch as a protective agent and β -D-glucose as a reductant in a mild-heating system. In this way, the attraction between starch and silver nanoparticles is weak and reversible at higher temperatures, facilitating the separation of the

synthesized silver nanoparticles (Mochochoko et al. 2013).

Tollens method

The Tollens method is carried out in a one-step process. In this approach, the reduction of Ag^+ ions is done by saccharides in the presence of ammonia, yielding silver nanoparticles with different shapes and sizes of 50–200 nm (Korbekandi and Iravani 2012). $\text{Ag}(\text{NH}_3)_2^+$ is a stable complex ion resulting from strong affinity of ammonia for Ag^+ ions, so the concentration of ammonia and nature of the reducing agents play a principal role in formulating the AgNP size (Dondi et al. 2012).

Irradiation method

Metal nanoparticles can be prepared by using various irradiation methods at room temperature without the use of reducing agent. Hence, temperature-dependent capping agents can also be used in the irradiation method. For instance, silver nanoparticles with a distinct shape and size distribution can be obtained from laser irradiation of an aqueous solution of silver salt and surfactant (Van Phu et al. 2014). Moreover, the morphology of metallic nanoparticles can be controlled by manipulating the radiation dose and dose rate (Abedini et al. 2013).

Biological method

Through biological method, extracts from biological agents such as microbes and plants can be employed either as reducing or protective agent for the fabrication of metal nanoparticles. In these extracts, various combinations of biomolecules which have the reducing potential can be found such as amino acids, vitamins, proteins, enzymes, and polysaccharides that are environmentally benign, yet chemically complex (Moghaddam 2010). For instance, the unicellular green algae *Chlorella vulgaris* extract was utilized to synthesize single-crystalline silver nano-plates at room temperature. Proteins in the extract were suggested to perform dual function of Ag^+ reduction and shape-control in the synthesis (Annamalai and Nallamuthu 2015).

Polyoxometalates method

Polyoxometalates are a vast family of molecular metal-oxide clusters with greater extent of structures. Meanwhile, their reduced forms possess greater capability of electron and proton transfer and/or storage abilities, and thus it can be employed to act as efficient donors or acceptors of several electrons without structural change. Hence, soluble polyoxometalates are capable of synthesizing noble nanoparticles through step-wise, multi-electron redox reactions inertly (Cauerhff and Castro 2013). For examples, a silver salt, Ag_2SO_4 , and polyoxometalates $(\text{NH}_4)_{10}[\text{Mo}^{\text{V}}]_4(\text{Mo}^{\text{VI}})_2\text{O}_{14}(\text{O}_3\text{PCH}_2\text{PO}_3)_2(\text{HO}_3\text{PCH}_2\text{PO}_3)_2 \cdot 15 \text{H}_2\text{O}$ and $\text{H}_7[\beta\text{-P}(\text{Mo}^{\text{VI}})_4(\text{Mo}^{\text{VI}})_8\text{O}_{40}]$ reacted to fabricate spherical and quasi-monodispersed silver nanoparticles with a diameter of about 38 nm after several minutes (Sharma et al. 2009).

In recent years, biological methods employing microbial organisms such as bacteria, actinomycetes, fungi, yeast, viruses, and also plants or plant extracts have gained considerable attention as an alternative to chemical and physical methods in the field of bio-nanotechnology (Khadri et al. 2013). As such, one of the fundamental processes in biosynthesis of nanoparticles involves bio-reduction. Many biological organisms, both unicellular and multicellular, have the ability to produce inorganic materials either intra- or extra-cellular, often of nanoscale dimensions and of exquisite morphology and hierarchical assembly (Pantidos and Horsfall 2014).

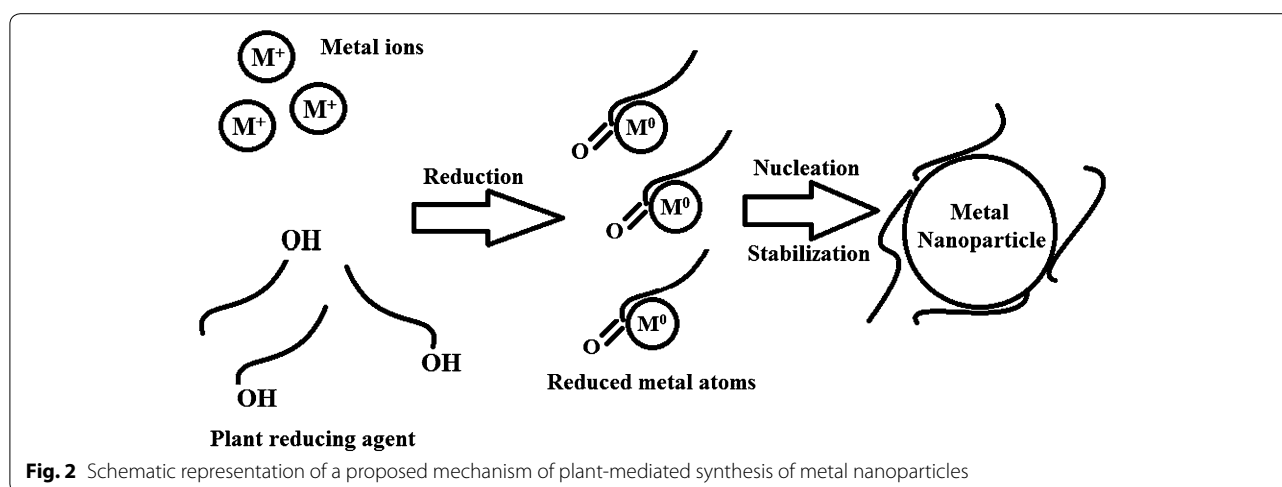
Plant-mediated synthesis of nanoparticles

Plant-mediated biosynthesis of nanoparticle is considered a widely acceptable technology for rapid production of metallic nanoparticles for successfully meeting the excessive need and current market demand and resulting in a reduction in the employment or generation of hazardous

substances to public health. Similar to microbes which have been used as a “bio-factory” in the synthesis of metallic nanoparticles, plants are also the natural “chemical factories” which are economical and require minimal maintenance (Nyoman Rupiasih et al. 2013).

Plants have several cellular structures and physiological processes to combat the toxicity of metals and maintain homeostasis. They also possess dynamic solutions to detoxify metals and hence scientists have now turned into phytoremediation (Abboud et al. 2013). The modus operandi of detoxification includes immobilization, exclusion, chelation, and compartmentalization of the metals ions, and the expression of more general stress response mechanisms, such as ethylene and stress proteins (Sánchez et al. 2011). The ability to tolerate inimical concentrations of toxic metals is found in the plant kingdom from ages. Their ability to accumulate high concentrations of metals was observed for both essential nutrients, such as copper (Cu), iron (Fe), zinc (Zn), and selenium, as well as non-essential metals, such as cadmium (Cd), mercury (Hg), lead (Pb), aluminum (Al), and arsenic (As) (Sahayaraj et al. 2012). In plants or plant-derived materials, a wide range of metabolites with redox potentials is determined, which are playing a principal role as a reducing agent in the biogenic synthesis of nanoparticles. In comparison to the microbial synthesis of nanoparticles, highly stable nanoparticles are synthesized by plant or plant extracts with the higher rate of production. Consequently, the advantages of plant-mediated preparation of metal nanoparticles lead researchers to in search of further exploration of the bio-reduction mechanism of metal ions by plants and the possible mechanism of formation of metal nanoparticle in and by the plants (Ahmad and Sharma 2012).

In recent years, biosynthesis of metal nanoparticles, especially silver and gold nanoparticles, using plant extracts as nano-factories becomes an important subject of researches in the field of bio-nanotechnology (Irvani 2011). Based on all aforementioned information, a schematic diagram of a proposed mechanism for plant-mediated fabrication of metal nanoparticles is illustrated in Fig. 2. Generally, the bio-reduction mechanism of metal nanoparticle in plants and plant extracts includes three main phases (Makarov et al. 2014). The activation phase in which the reduction of metal ions and nucleation of the reduced metal atoms occur. The growth phase, referring to the spontaneous coalescence of the small adjacent nanoparticles into particles of a larger size, accompanied by an increase in the thermodynamic stability of nanoparticles, or a process referred to as Ostwald ripening and the termination phase in which the final shape of the nanoparticles formed.



Advantages of plant-mediated synthesis of nanoparticles

Due to their easy availability, green preparation of nanoparticles using plant extracts turns out to be an important research subject in the field of bio-nanotechnology in this era. Principally, the biogenic synthesis employs plant extracts in aqueous form in the fabrication of noble nanoparticles for the reason that the availability of reducing agent is higher in the extract than the whole plant (Huang et al. 2007). Besides, plant-mediated synthesis of nanoparticles is simpler and easier to be conducted without requiring any specific operating conditions as compared to typical physical and chemical methods. The synthesized products of the process including waste products are resulted from natural plant extracts, and hence this technique is also more environmental green. Nevertheless, both strong and weak chemical reducing agents and capping agents such as sodium citrate, sodium borohydride, and alcohols, which are mostly toxic, flammable, and cannot be degraded easily are required in the physical and chemical methods (Lalitha et al. 2013).

Through this bio-based protocol of nanoparticles synthesis, higher reproducibility of the process and higher stability of the synthesized nanoparticles can be attained. Therefore, this green-based fabrication of nanoparticles is suitable for large scale production with more effective cost investment, eco-friendly, and safe for human therapeutic use. Apart from the aspects of reproducibility and stability, the rate of bio-reduction of metal ions using biological agents is showed to be much faster and also at ambient temperature and pressure conditions (Pasupuleti et al. 2013). On the contrary, previous studies reported that the bio-reduction potential of the plant extracts is comparatively higher than the microbial culture (Khalil et al. 2014). Moreover, the waste products resulted from the microbial-based method is likely to

be more harmful to the environment depending on the type of microbes involved in the synthesis (Moghaddam 2010). Hence, plant-mediated synthesis brings less or almost zero contamination and so reducing the impact on the environment. With all the aforementioned advantages and outstanding features over other methods, the biosynthetic method employing plant extracts has now turned as a simple, effective and viable technique as well as a good alternative to conventional chemical and physical nanoparticle preparation methods, and even microbial methods (Huang et al. 2007).

Silver nanoparticles

Silver is a gleaming, very ductile, and malleable element but slightly harder than gold, with a symbol of Ag and atomic number of 47. It is one of the basic elements that make up our planet. In nature, it exists as a native element, as an alloy combining with other metals (e.g., gold) and as minerals (e.g., chlorargyrite and argentite). Chemically, silver possess four different oxidation states, i.e., Ag^0 , Ag^{1+} , Ag^{2+} , and Ag^{3+} (Riedel and Kaupp 2009). However, it is a chemically inactive element, but it can be reacted with nitric acid or hot concentrated sulfuric acid, forming soluble silver salts. It also possesses an excellent conductivity of heat and electricity, yet its applications in electrical industry have greatly been limited due to its greater cost (Wang et al. 2013). As for metallic silver form, it is insoluble in water, but its metallic salts such as silver nitrate, $AgNO_3$, and silver chloride, $AgCl$, are water-soluble. Over past decades, metallic silver is widely applied in surgical prosthesis and splints, coinage, and fungicides (Forough and Farhadi 2010). In contrast, its metallic salts have also been made use of treating various health illness and disorders, e.g., epilepsy, gonorrhoea, and gastroenteritis. Due to its good absorptivity, soluble

silver compounds have the risk of causing adverse effects on health through dietary intake. Nevertheless, Chen and Schluesener (2008) described that silver is relatively non-toxic and non-carcinogenic to human primary body systems such as nervous, immune, reproductive, or cardiovascular system. Therefore, the demand of silver has been escalating in recent years, exclusively in medical, plastics, and textiles industries.

Since silver is non-toxic to animal cells, it has been considered as a safe and effective anti-bactericidal metal, especially it is highly toxic to bacteria such as *Escherichia coli* (*E. coli*) and *Staphylococcus aureus* (El-Kheshen and El-Rab 2012). Hence, silver-based compounds have received much attention as an antimicrobial agent for centuries to prevent bacterial growth in applications such as burn care (Pasupuleti et al. 2013). In recent years, due to the increasing threat of antibiotic resistance that is caused by the abuse of antibiotics, it has been a driving force leading to in search for the synthesis of silver nanoparticles (Table 1).

Antimicrobial properties of silver nanoparticles

The exact mechanism of antibacterial activity of silver nanoparticles on microbes is yet to be evaluated, but it can be linked to the mechanism of Ag^+ ions action against bacteria stains such as trypanosomes and yeasts, whereby the occurrence of buildup of AgNPs from the aqueous solution eventually causes saturation of enzymes and protein in the cell. Instead, there are three possible antibacterial mechanisms of silver nanoparticles that have been proposed by Li and coworkers (Li et al. 2008):

- (a) Bacterial growth and proliferation are adversely inhibited by the adhesion of ultra-small sized silver nanoparticles onto the cell wall of bacteria, resulting in changes in the cell wall which in turn is unable to protect the interior of the cell;
- (b) Through the penetration of silver nanoparticles into the bacterial cell, it leads to DNA damage, or even cell death, by altering its normal functioning of bacterial DNA; and
- (c) The interaction of Ag^+ ions with the proteins containing sulfur present in the bacterial cell wall irreversibly caused the disruption of the bacterial cell wall. This proposed mechanism is also deduced as the main antibacterial mechanism in evaluating the antimicrobial activity.

The antimicrobial effect of silver nanoparticles depends on various parameters including size, shape, and the surface charge of the particles. In this respect, nanoparticles have greater antibacterial properties as they can easily penetrate into the nuclear content of bacteria due to their

structure of the bacterial cell wall, especially in gram-negative bacteria, inactivating DNA and the enzymes leading to cellular death. They can also possess a greater surface area for stronger bactericidal interactions (Pal et al. 2007). Furthermore, the antibacterial activity of silver nanoparticles also depends on the morphology of the nanoparticles. Pal et al. (2007) reported that silver nanoparticles with the same surface areas but with different shapes can exhibit dissimilar antibacterial activity which might be due to the difference in their effective surface areas and number of active facets. Truncated triangular silver nano-plates that were found to display the strongest antibacterial activity could be due to their larger surface area to volume ratios and their crystallographic surface structures. In addition, the electrostatic attraction between positively charged nanoparticles and negatively charged bacterial cells is another important factor contributing to the antimicrobial activity of silver nanoparticles. In gram-negative bacteria such as *Escherichia coli*, *Pseudomonas*, *Salmonella*, and *Vibrio*, its cell wall consists of a layer of lipopolysaccharide at the external surface followed by a thin layer of peptidoglycan. As comparison, the cell wall in gram-positive bacteria such as *Bacillus*, *Clostridium*, *Staphylococcus*, and *Streptococcus*, is mainly composed of a thick layer of peptidoglycan (Morones et al. 2005). Although gram-positive and gram-negative bacteria have differences in their membrane structures, most of them possess a negative charge on their surfaces. Hence, silver nanoparticles exhibit greater antimicrobial effect against gram-negative bacteria regardless of their resistance level as compared to gram-positive bacteria (Abbaszadegan et al. 2015).

Application of silver nanoparticles

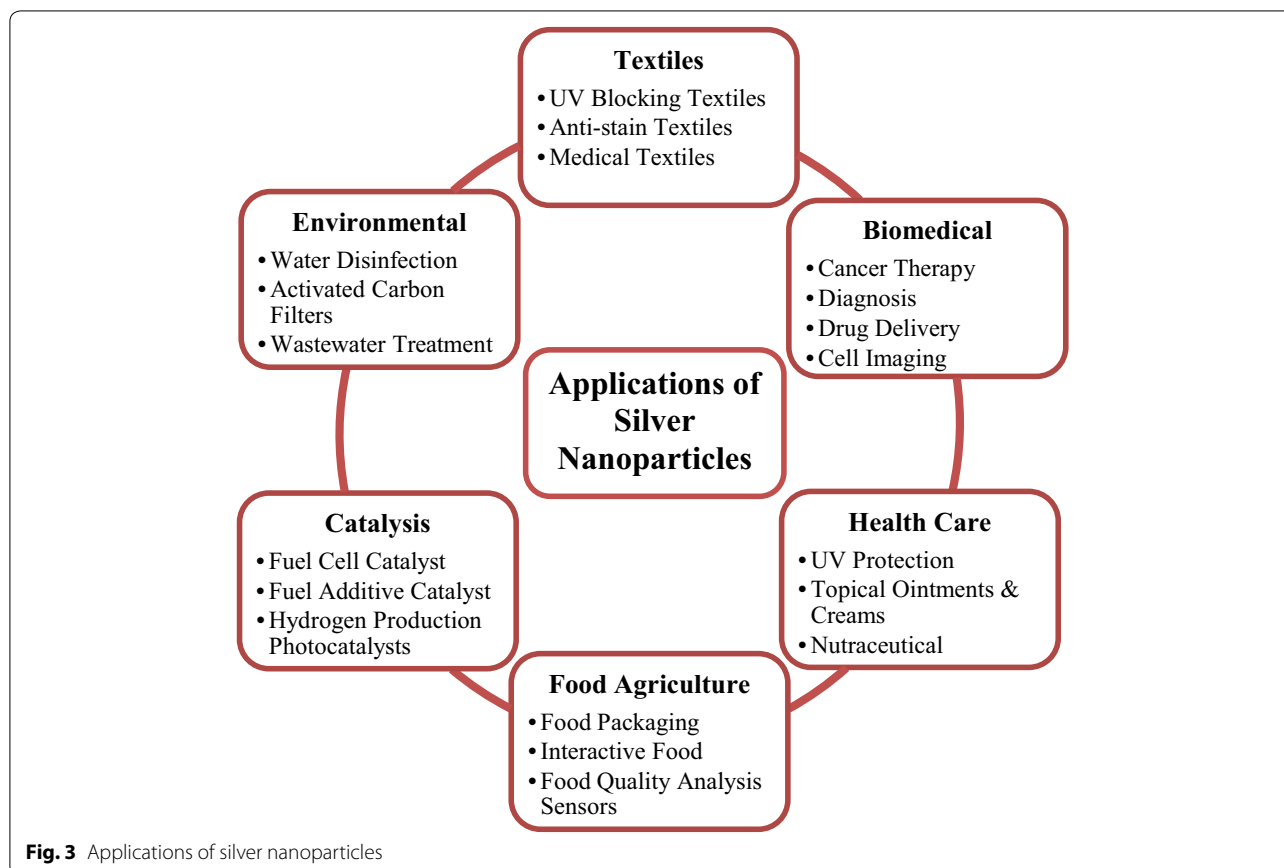
Silver nanoparticles are of particular interest in the modern research of nanotechnology due to its unique properties, which can be incorporated into a wide range of extensive applications such as antiseptic agents in medical industry, cosmetics, food packaging, bioengineering, electrochemistry, catalysis, and environmental uses. As compared to their bulk materials, noble nanoparticles exhibit different catalytic activities, and therefore nanocatalysis recently has gained much attention in such a way of using nanoparticles as catalysts in various types of processes. For instance, gold, silver, platinum, and metal ions are well-known catalysts in the process of decomposition of H_2O_2 to oxygen (Merga et al. 2007). In the study of Guo et al. (2008), catalytic potential of silver nanoparticles was observed to be better than that of gold and platinum nanoparticles in the emission system of chemiluminescence from luminol- H_2O_2 . Moreover, catalysis of the reduction of dyes by sodium borohydride (NaBH_4) can be enhanced by using silver nanoparticles

immobilized on silica spheres. In the absence of silver nanoparticles as catalysts, the rate of reaction was almost stationary showing very little or even no reduction of the dyes occurred (Guo et al. 2008) (Fig. 3).

Due to the significant antimicrobial properties of AgNPs against wide ranging microorganisms, numerous medical applications impregnated with AgNPs such as catheters and cardiovascular and bone implants have been recognized for hindering the formation of biofilm and lowering the risk of pathogenic invasion (Tran et al. 2013). Typically, ultrahigh molecular weight polyethylene has been widely used as an insert for artificial joint replacement, but its application is somehow limited due to its high susceptibility to wear and tear (Morley et al. 2007). Yet, the drawback of wear and tear of the polymer is significantly abridged by the addition of silver nanoparticles. In addition, with the excellent antibacterial properties of silver nanoparticles, it is also loaded with polymethyl methacrylate broadly as bone cements that are broadly used as synthetic joint replacement (Alt et al. 2004). In 2010, Xing et al. deduced that (poly(-3-hydroxybutyrate-co-3-hydroxyvalerate) PHBV nanofiber scaffolds containing AgNPs have the tendency of aiding in bone and skin tissue regeneration from their extensive

study on both osteoblast (bone cells) and fibroblast (skin cells) cultured on such scaffolds. Hence, the risk associated with implantation surgery can be overcome by fabricating the surface of structure of the bone implants devices and scaffolds with silver nanoparticles (Xing et al. 2010).

Furthermore, silver nanoparticles are also applied in nano-crystalline dressings for the therapy of wound or hospital-acquired infections, minimizing the inflammatory response (Fong and Wood 2006). For instance, the classical surgical meshes are used to bridge severe wounds and for tissue therapy, but it is highly vulnerable to pathogenic invasions. Thus, the effectiveness of these meshes is enhanced with the impregnation with silver nanoparticles. With the plasmonic properties of silver nanoparticles, it can be widely applied in bio-imaging for monitoring dynamic events over an extended period of time without undergoing photo-bleaching as compared to common fluorescent dyes. Therefore, the conjugation of cells to the target cells leads to the conversion of light energy to thermal energy and then resulted in thermal ablation of the target cells, aiding in destroying unwanted or damaged cells (Loo et al. 2005). In addition, the plasmonic properties of silver nanoparticles can be exploited



for bio-sensing, which can effectively detect wide ranging of proteins that typical biosensors do not. With this unique capability, silver nanoparticles are broadly employed for detecting various abnormalities and diseases in human body system, e.g., tumor cells or cancer. The plasmonic properties of silver nanoparticles are somehow dependent on its size, shape, and the dielectric potential of surrounding medium (Morley et al. 2007).

In recent years, silver nanoparticles are widely applied in chemical industry as an additive to cosmetics, because silver nanoparticles satisfy the requirements of excellent antiseptic properties, as a safe preservative additive, and also as a constituent for the skin therapy, e.g., treatment of acne (Kokura et al. 2010). In addition to the applications of silver nanoparticles in medical and environmental protection field, silver nanoparticle-coated paper could also serve a critical role in food preservation in which provides a reservoir for slow releasing of ionic silver from the surface to the bulk to prevent microbial growth in the food as well as to prevent growth of pathogens on the surface itself (Gottesman et al. 2011). Owing to the excellent antimicrobial activity of silver nanoparticles, developing antibacterial coatings on surfaces has drawn much interest for human health and environmental protection in the paint coating industry. In 2008, John et al. demonstrated green synthesis techniques of metallic nanoparticle-embedded paints using common household paint in a single step. Through the naturally occurring oxidative drying process in oils that involves free-radical exchange, reduction of metal salts and dispersion of metal nanoparticles in the oil media were successfully done without the use of any external chemical reducing or stabilizing agents. The resulting well-dispersed metal nanoparticles in oil dispersions can then be directly used on different surfaces such as wood, glass, steel, and different polymer surfaces, and also exhibit excellent bactericidal properties against gram-positive and gram-negative bacteria, especially silver nanoparticle-embedded paints (Kumar et al. 2008).

Toxicity of silver nanoparticles

Generally, silver nanoparticles can be considered as an ideal candidate for numerous applications in various fields, especially in biomedical industry as in diagnosis, drug delivery, cell imaging, and implantation, even so several studies reported that silver nanoparticles possess an adverse effect on humans as well as the environment. In one of the toxicology researches of silver nanoparticles, *in vitro* toxicity assay in rat liver cells has conducted and demonstrated that silver nanoparticles caused oxidative stress and cease of mitochondrial function even at low level of exposure to silver nanoparticles (10–50 $\mu\text{g ml}^{-1}$). Yet, at higher doses (>1.0 mg L^{-1}), AgNPs

exhibited a significant cytotoxicity and caused abnormal cellular morphology, cellular shrinkage, and acquisition of an irregular shape (McAuliffe and Perry 2007). Besides, silver nanoparticles can also induce toxicity to *in vitro* mouse germ line stem cells by impairing mitochondrial activities and cause leakage through the cell membranes by changing its permeability to sodium and potassium ions. Therefore, the cytotoxic mechanism of AgNPs is predominantly based on the induction of reactive oxygen species (ROS). Specifically, exposure to silver nanoparticles triggers depletion in glutathione level, elevation of ROS levels, lipid peroxidation, and increased expression of ROS responsive genes, leading to DNA damage, apoptosis, and necrosis (Haider and Kang 2015).

As a result of large surface area to volume ratio, human bodies are vulnerably exposed to silver nanoparticles via ingestion, inhalation, or skin, and its penetrating potential is greatly increased, and hence it has the capability of penetrating into the circulatory system and even translocating boundlessly in the human body system (Sung et al. 2008). Hence, previous research elaborated that silver nanoparticles can allegedly induce toxicity to the male reproductive system by crossing through blood-testes barrier and depositing in the testes. Moreover, silver nanoparticles can induce adversely toxic effect on the proliferation and cytokine expression by peripheral blood mononuclear cells (McAuliffe and Perry 2007). Alternatively, Kim et al. evaluated that silver nanoparticles do not induce genetic toxicity in male and female Sprague–Dawley rat bone marrow *in vivo* in his gastrointestinal toxicology study. In his experiment, there were no significant changes in body weight of male and female rats relative to the doses of AgNPs (size of 60 nm) over a period of 28 days. However, alkaline phosphatase and cholesterol values were found to be altered with the exposure to over more than 300 mg of AgNPs, resulting in minor liver damage (Kim et al. 2008).

Though aforementioned studies tend to suggest that silver nanoparticles can adversely induce toxicity to living beings, relatively less *in vivo* toxicology researches of silver nanoparticles were established which are drastically different from *in vitro* condition. Thus, further investigation is required to assess the toxicity effect of silver nanoparticles in *in vivo* condition for evaluating its exact toxicity to human and animals.

Conclusion

Chemical and physical syntheses of nanoparticles are unable to be expanded easily to large scale production due to several drawbacks such as the presence of toxic organic solvents, production of hazardous by-products and intermediate compounds, and high energy consumption. This could lead to an increase in the particle

reactivity and toxicity, which might harm human health and environment due to the composition ambiguity and lack of predictability. Therefore, this leads to biological methods which could be more eco-friendly and does not cause any harm to human and domestic animals health.

Authors' contributions

NAE was responsible for the manuscript approval and final approval. AME was responsible for manuscript writing; AA and CLK were responsible for manuscript drafting. All authors read and approved the final manuscript.

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Competing interests

The authors of this manuscript do not have any financial competing interests to declare.

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References

- Abbaszadegan A, Ghahramani Y, Gholami A, Hemmateenejad B, Dorostkar S, Nabavizadeh M, Sharghi H (2015) The effect of charge at the surface of silver nanoparticles on antimicrobial activity against gram-positive and gram-negative bacteria: a preliminary study. *J Nanomat*
- Abboud Y, Eddahbi A, El Bouari A, Aitenneite H, Brouzi K, Mouslim J (2013) Microwave-assisted approach for rapid and green phyto-synthesis of silver nanoparticles using aqueous onion (*Allium cepa*) extract and their antibacterial activity. *J Nanostruct Chem* 3(1):1–7. doi:10.1186/2193-8865-3-84
- Abedini A, Daud AR, Hamid MAA, Othman NK, Saion E (2013) A review on radiation-induced nucleation and growth of colloidal metallic nanoparticles. *Nanoscale Res Lett* 8(1):1–10
- Abou El-Nour KMM, Aa Eftaiha, Al-Warthan A, Ammar RAA (2010) Synthesis and applications of silver nanoparticles. *Arab J Chem* 3(3):135–140. doi:10.1016/j.arabj.2010.04.008
- Adlakh-Hutcheon G, Khaydarov R, Korenstein R, Varma R, Vaseashta A, Stamm H, Abdel-Mottaleb M (2009) Nanomaterials, nanotechnology. In: Linkov I, Steevens J (eds) *Nanomaterials: Risks and Benefits*. NATO Science for Peace and Security Series C: Environmental Security. Springer, Netherlands, pp 195–207. doi:10.1007/978-1-4020-9491-0_14
- Ahmad Naheed, Sharma S (2012) Green synthesis of silver nanoparticles using extracts of *Ananas comosus*. *Green Sustain Chem* 2:141–147
- Alt V, Bechert T, Steinrücke P, Wagener M, Seidel P, Dingeldein E, Domann E, Schnettler R (2004) An in vitro assessment of the antibacterial properties and cytotoxicity of nanoparticulate silver bone cement. *Biomaterials* 25(18):4383–4391
- Annamalai J, Nallamuthu T (2015) Green synthesis of silver nanoparticles: characterization and determination of antibacterial potency. *Appl Nanosci* 1–7
- Arun G, Eyini M, Gunasekaran P (2014) Green synthesis of silver nanoparticles using the mushroom fungus *Schizophyllum commune* and its biomedical applications. *Biotechnol Bioprocess Eng* 19(6):1083–1090. doi:10.1007/s12257-014-0071-z
- Awwad A, Salem N, Abdeen A (2013) Green synthesis of silver nanoparticles using carob leaf extract and its antibacterial activity. *Int J Ind Chem* 4(1):1–6. doi:10.1186/2228-5547-4-29
- Azizi S, Namvar F, Mahdavi M, Ahmad MB, Mohamad R (2013) Biosynthesis of silver nanoparticles using brown marine macroalgae, *Sargassum muticum* aqueous extract. *Materials*. doi:10.3390/ma6125942
- Banerjee P, Satapathy M, Mukhopahayay A, Das P (2014) Leaf extract mediated green synthesis of silver nanoparticles from widely available Indian plants: synthesis, characterization, antimicrobial property and toxicity analysis. *Bioresour Bioprocess* 1(1):3
- Birla SS, Gaikwad SC, Gade AK, Rai MK (2013) Rapid synthesis of silver nanoparticles from *Fusarium oxysporum* by optimizing physiocultural conditions. *Scient World J* 2013:12. doi:10.1155/2013/796018
- Cauerhff Ana, Castro GR (2013) Bionanoparticles, a green nanochemistry approach. *Electron J Biotechnol*. doi:10.2225/vol16-issue3-fulltext-3
- Chaturvedi V, Verma P (2015) Fabrication of silver nanoparticles from leaf extract of *Butea monosperma* (Flame of Forest) and their inhibitory effect on bloom-forming cyanobacteria. *Bioresour Bioprocess* 2(1):18
- Chen X, Schluesener HJ (2008) Nanosilver: a nanoproduct in medical application. *Toxicol Lett* 176(1):1–12. doi:10.1016/j.toxlet.2007.10.004
- Dondi R, Su W, Griffith GA, Clark G, Burley GA (2012) Highly size- and shape-controlled synthesis of silver nanoparticles via a templated tollens reaction. *Small* 8(5):770–776
- Durán Nelson, Marcato Priscyla D, De Conti Roseli, Alves Oswaldo L, Costab FTM, Brocchib M (2010) Potential use of silver nanoparticles on pathogenic bacteria, their toxicity and possible mechanisms of action. *J Braz Chem Soc* 21(6):949–959
- Edhaya Naveena B, Prakash S (2013) Biological synthesis of gold nanoparticles using marine algae *Gracilaria corticata* and its application as a potent antimicrobial and antioxidant agent. *Asian J Pharm Clin Res* 6(2):179–182
- El-Batal AI, Amin MA, Shehata MM, Hallol MM (2013) Synthesis of silver nanoparticles by *Bacillus stearothermophilus* using gamma radiation and their antimicrobial activity. *World Appl Sci J* 22(1):01–16. doi:10.5829
- El-Kheshen Amany A, El-Rab SFG (2012) Effect of reducing and protecting agents on size of silver nanoparticles and their anti-bacterial activity. *Schol Res Librar* 4(1):53–65
- El-Shishtawy RM, Asiri AM, Al-Otaibi MM (2011) Synthesis and spectroscopic studies of stable aqueous dispersion of silver nanoparticles. *Spectrochim Acta Part A Mol Biomol Spectrosc* 79(5):1505–1510. doi:10.1016/j.saa.2011.05.007
- EU (2011) Commission Recommendation of 18 October 2011 on the definition of nanomaterial (2011/696/EU). Official Journal of the European Union 2011 L275/38
- Fong J, Wood F (2006) Nanocrystalline silver dressings in wound management: a review. *Int J Nanomed* 1(4):441–449
- Forough Mehrdad, Farhadi K (2010) Biological and green synthesis of silver nanoparticles. *Turkish J Eng Env Sci* 34:281–287. doi:10.3906/muh-1005-30
- Gottesman R, Shukla S, Perkas N, Solovyov LA, Nitzan Y, Gedanken A (2011) Sonochemical coating of paper by microbicidal silver nanoparticles. *Langmuir* 27(2):720–726. doi:10.1021/la103401z
- Guo J-Z, Cui H, Zhou W, Wang W (2008) Ag nanoparticle-catalyzed chemiluminescence reaction between luminol and hydrogen peroxide. *J Photochem Photobiol A* 193(2–3):89–96
- Haider A, Kang H-K (2015) Preparation of silver nanoparticles and their industrial and biomedical applications: a comprehensive review. *Adv Mat Sci Eng*
- Heydari R, Rashidipour M (2015) Green synthesis of silver nanoparticles using extract of oak fruit hull (Jaft): synthesis and in vitro cytotoxic effect on MCF-7 cells. *Int J Breast Cancer* 2015:6. doi:10.1155/2015/846743
- Huang Jiale, Qingbiao L, Sun Daohua, Yinghua Lu, Yuanbo Su, Yang Xin, Wang Huixuan, Wang Yuaneng, Shao Wenyao, He Ning, Hong Jinqing, Chen C (2007) Biosynthesis of silver and gold nanoparticles by novel sundried *Cinnamomum camphora* leaf. *Nanotechnology*. doi:10.1088/0957-4484/18/10/105104
- Iravani S (2011) Green synthesis of metal nanoparticles using plants. *Green Chem* 13(10):2638–2650. doi:10.1039/C1GC15386B
- Johnson I, Prabu HJ (2015) Green synthesis and characterization of silver nanoparticles by leaf extracts of *Cycas circinalis*, *Ficus amplissima*, *Commelina benghalensis* and *Lippia nodiflora*. *Int Nano Lett* 5(1):43–51. doi:10.1007/s40089-014-0136-1
- Karthik L, Kumar G, Kirthi AV, Rahuman AA, Bhaskara Rao KV (2014) *Streptomyces* sp. LK3 mediated synthesis of silver nanoparticles and its biomedical application. *Bioprocess Biosyst Eng* 37(2):261–267. doi:10.1007/s00449-013-0994-3
- Kathiresan K, Manivannan S, Nabeel MA, Dhivya B (2009) Studies on silver nanoparticles synthesized by a marine fungus, *Penicillium fellutanum* isolated from coastal mangrove sediment. *Colloids Surf B* 71(1):133–137. doi:10.1016/j.colsurfb.2009.01.016

- Khabat Vahabi G, Mansoori Ali, Karimi S (2011) Biosynthesis of silver nanoparticles by fungus *Trichoderma Reesei*. *Insci J* 1(1):65–79. doi:10.5640/insc.010165
- Khadri Habeeb, Alzohary Mohammad, Janardhan Avilala, Kumar Arthala Praveen, Narasimha G (2013) Green synthesis of silver nanoparticles with high fungicidal activity from olive seed extract. *Adv Nanopart* 2:241–246
- Khalil MMH, Ismail EH, El-Baghady KZ, Mohamed D (2014) Green synthesis of silver nanoparticles using olive leaf extract and its antibacterial activity. *Arab J Chem* 7(6):1131–1139. doi:10.1016/j.arabj.2013.04.007
- Khatami M, Pourseyedi S, Khatami M, Hamidi H, Zaeifi M, Soltani L (2015) Synthesis of silver nanoparticles using seed exudates of *Sinapis arvensis* as a novel bioresource, and evaluation of their antifungal activity. *Bioresour Bioprocess* 2(1):19
- Kim YS, Kim JS, Cho HS, Rha DS, Kim JM, Park JD, Choi BS, Lim R, Chang HK, Chung YH (2008) Twenty-eight-day oral toxicity, genotoxicity, and gender-related tissue distribution of silver nanoparticles in Sprague-Dawley rats. *Inhalation Toxicol* 20(6):575–583
- Kokura Satoshi, Handa Osamu, Takagi Tomohisa, Ishikawa Takeshi, Naito Yuji, Yoshikawa T (2010) Silver nanoparticles as a safe preservative for use in cosmetics. *Nanotechnology: Biol Med* 6:570–574. doi:10.1016/j.nano.2009.12.002
- Korbekandi H, Iravani S (2012) Silver Nanoparticles. INTECH Open Access Publisher
- Kumar A, Vemula PK, Ajayan PM, John G (2008) Silver-nanoparticle-embedded antimicrobial paints based on vegetable oil. *Nat Mater* 7(3):236–241. doi:10.1038/nmat2099
- Kumar B, Kumari S, Cumbal L, Debut A (2015) *Lantana camara* berry for the synthesis of silver nanoparticles. *Asian Pacific J Trop Biomed* 5(3):192–195. doi:10.1016/S2221-1691(15)30005-8
- Lalitha A, Subbaiya R, Ponmurugan P (2013) Green synthesis of silver nanoparticles from leaf extract *Azadirachta indica* and to study its anti-bacterial and antioxidant property. *Int Curr Microbiol Appl Sci* 2(6):228–235
- Li Q, Mahendra S, Lyon DY, Brunet L, Liga MV, Li D, Alvarez PJ (2008) Antimicrobial nanomaterials for water disinfection and microbial control: potential applications and implications. *Water Res* 42(18):4591–4602
- Loo C, Lowery A, Halas N, West J, Drezek R (2005) Immuno targeted nanoshells for integrated cancer imaging and therapy. *Nano Lett* 5(4):709–711
- Madhuri S, Maheshwar S, Sunil P, Oza G (2012) Nanotechnology: concepts and applications, vol 4. CRC Press, USA
- Makarov VV, Love AJ, Sinitysyna OV, Makarova SS, Yaminsky IV, Taliany ME, Kalinina NO (2014) “Green” nanotechnologies: synthesis of metal nanoparticles using plants. *Acta Naturae* 6(1):35–44
- Malik P, Shankar R, Malik V, Sharma N, Mukherjee TK (2014) Green chemistry based benign routes for nanoparticle synthesis. *J Nanopart* 2014:14. doi:10.1155/2014/302429
- Mason Cynthis, Vivekanadhan Singaravelu, Misra Manjusri, Mohanty AK (2012) Switchgrass (*Panicum virgatum*) extract mediated green synthesis of silver nanoparticles. *World J Nano Sci Eng* 2:47–52
- McAuliffe ME, Perry MJ (2007) Are nanoparticles potential male reproductive toxicants? A literature review. *Nanotoxicology* 1(3):204–210
- Méndez-Vilas A (2011) Science Against Microbial Pathogens: Communicating Current Research and Technological Advances. vol v. 2. Formatex Research Center
- Merga G, Wilson R, Lynn G, Milosavljevic BH, Meisel D (2007) Redox catalysis on “Naked” silver nanoparticles. *J Phys Chem C* 111(33):12220–12226. doi:10.1021/jp074257w
- Metuku R, Pabba S, Burra S, Hima Bindu N SVSSSL, Gudikandula K, Singara Charya MA (2014) Biosynthesis of silver nanoparticles from *Schizophyllum radiatum* HE 863742.1: their characterization and antimicrobial activity. *Biotech* 4(3):227–234. doi:10.1007/s13205-013-0138-0
- Mochochoko T, Oluwafemi OS, Jumbam DN, Songca SP (2013) Green synthesis of silver nanoparticles using cellulose extracted from an aquatic weed; water hyacinth. *Carbohydr Polym* 98(1):290–294. doi:10.1016/j.carbpol.2013.05.038
- Moghaddam KM (2010) An Introduction to microbial metal nanoparticle preparation method. *J Young Invest* 19(19):1–6
- Mohammed AE (2015) Green synthesis, antimicrobial and cytotoxic effects of silver nanoparticles mediated by *Eucalyptus camaldulensis* leaf extract. *Asian Pacific J Trop Biomed* 5(5):382–386. doi:10.1016/S2221-1691(15)30373-7
- Mohanpuria P, Rana N, Yadav S (2008) Biosynthesis of nanoparticles: technological concepts and future applications. *J Nanopart Res* 10(3):507–517. doi:10.1007/s11051-007-9275-x
- Morley K, Webb P, Tokareva N, Krasnov A, Popov V, Zhang J, Roberts C, Howdle S (2007) Synthesis and characterisation of advanced UHM-WPE/silver nanocomposites for biomedical applications. *Eur Polymer J* 43(2):307–314
- Morones JR, Elechiguerra JL, Camacho A, Holt K, Kouri JB, Ramírez JT, Yacaman MJ (2005) The bactericidal effect of silver nanoparticles. *Nanotechnology* 16(10):2346
- Nithya Deva Krupa A, Raghavan V (2014) Biosynthesis of silver nanoparticles using *Aegle marmelos* (Bael) fruit extract and its application to prevent adhesion of bacteria: a strategy to control microfouling. *Bioinorg Chem Appl* 2014:8. doi:10.1155/2014/949538
- Nyoman Rupiasih N, Aher Avinash, Gosavi Suresh, Vidyasagar PB (2013) Green synthesis of silver nanoparticles using latex extract of *Thevetia peruviana*: a novel approach towards poisonous plant utilization. *J Phys Conf Series*. doi:10.1088/1742-6596/423/1/012032
- Oei JD, Zhao WW, Chu L, DeSilva MN, Ghimire A, Rawls HR, Whang K (2012) Antimicrobial acrylic materials with in situ generated silver nanoparticles. *J Biomed Mater Res B Appl Biomater* 100(2):409–415. doi:10.1002/jbm.b.31963
- Pal S, Tak YK, Song JM (2007) Does the antibacterial activity of silver nanoparticles depend on the shape of the nanoparticle? A study of the gram-negative bacterium *Escherichia coli*. *Appl Environ Microbiol* 73(6):1712–1720
- Pal Sovan Lal, Utpal Jana PK, Manna GP, Mohanta Manavalan R (2011) Nanoparticle: an overview of preparation and characterization. *J Appl Pharma Sci* 1(6):228–234
- Pantidos Nikolaos, Horsfall LE (2014) Biological synthesis of metallic nanoparticles by bacteria, fungi and plants. *Nanomed Nanotechnol*. doi:10.4172/2157-7439.1000233
- Pasupuleti VR, Prasad T, Shiekh RA, Balam SK, Narasimhulu G, Reddy CS, Rahman IA, Gan SH (2013) Biogenic silver nanoparticles using *Rhinacanthus nasutus* leaf extract: synthesis, spectral analysis, and antimicrobial studies. *Int J Nanomed* 8:3355–3364. doi:10.2147/IJN.S49000
- Prabhu S, Poulouse E (2012) Silver nanoparticles: mechanism of antimicrobial action, synthesis, medical applications, and toxicity effects. *Int Nano Lett* 2(1):1–10. doi:10.1186/2228-5326-2-32
- Qi WH, Wang MP (2004) Size and shape dependent melting temperature of metallic nanoparticles. *Mater Chem Phys* 88:280–284. doi:10.1016/j.matchemphys.2004.04.026
- Raffin M, Hussain F, Bhatti TM, Akhter JI, Hameed A, Hasan MM (2008) *J Mat Sci Technol* 24:192–196
- Rajathi K, Vijaya Raj D, Anarkali J, Sridhar S (2012) Green Synthesis, characterization and in-vitro antibacterial activity of silver nanoparticles by using *Tinospora cordifolia* leaf extract. *Int J Green Chem Bioprocess* 2(2):15–19
- Raut Rajesh W, Lakkakula Jaya R, Kolekar Niranjan S, Mendhulkar Vijay D, Kashid Sahebrao B (2009) Photosynthesis of Silver Nanoparticle Using *Gliricidia sepium* (Jacq.). *Current Nanoscience* 5(1):117–122
- Riedel Sebastian, Kaupp Martin (2009) The highest oxidation states of the transition metal elements. *Coord Chem Rev* 253(5–6):606–624. doi:10.1016/j.ccr.2008.07.014
- Roduner E (2006) Size matters: why nanomaterials are different. *Chem Soc Rev* 35(7):583–592. doi:10.1039/B502142C
- Sahayaraj K, Rajesh S, Rathil JM (2012) Silver nanoparticles biosynthesis using marine alga *Padina pavonica* (Linn.) and its microbicidal activity. *Digest J Nanomat Biostruct* 7(4):1557–1567
- Saikia Dulen, Gogoi Pradip K, Phukan Pallabi, Bhuyan Nilave, Borchetia Sang-eeta, Saika J (2015) Green synthesis of silver nanoparticles using *Asiatic Pennywort* and *Bryophyllum* leaves extract and their antimicrobial activity. *Adv Mat Lett* 6(3):260–264. doi:10.518/amlett.2015.5655
- Sánchez Eldidio Morales-, Guahardo-Pacheco Jesús, Noriega-Treviño María, Quintero-González Cristina, Compeán-Jasso Martha, López-Salinas Francisco, González-Hernández Jesús, Ruiz F (2011) Synthesis of silver nanoparticles using albumin as a reducing agent. *Mat Sci Appl* 2:578–581. doi:10.4236/msa.2011.26077
- Sarahlbrahim H, Ayad M, Fadhil A, Nk Al-Ani (2014) Production of Ag nanoparticles using *Aloe vera* extract and its antimicrobial activity. *J Al-Nahrain Univ* 17(2):165–171

- Saraniya Devi J, Valentin Bhimba B, Peter DM (2013) Production of biogenic Silver nanoparticles using *Sargassum longifolium* and its application. *Indian J Geo-Marine Sci* 42(7):125–130
- Sasikala D, Govindaraju K, Tamilselvan S, Singaravelu G (2012) Soybean protein: a natural source for the production of green silver nanoparticles. *Biotechnol Bioprocess Eng* 17(6):1176–1181. doi:10.1007/s12257-012-0021-6
- Shams Somaye, Pourseyedi Shahram, Raisi M (2013) Green Synthesis of Ag nanoparticles in the present of *Lens culinaris* seed exudates. *Int J Agricul Crop Sci* 5(23):2812–2815
- Shanmugavadivu M, Selvam K, Ranjithkumar R (2014) Synthesis of pomegranate peel extract mediated silver nanoparticles and its antibacterial activity. *Am J Adv Drug Deliv* 2(2)
- Sharma VK, Yngard RA, Lin Y (2009) Silver nanoparticles: green synthesis and their antimicrobial activities. *Adv Colloid Interface Sci* 145(1):83–96
- Sung JH, Ji JH, Yoon JU, Kim DS, Song MY, Jeong J, Han BS, Han JH, Chung YH, Kim J (2008) Lung function changes in Sprague-Dawley rats after prolonged inhalation exposure to silver nanoparticles. *Inhalation Toxicol* 20(6):567–574
- Tran Quang Huy, Nguyenm VQ, Le A-T (2013) Silver nanoparticles: synthesis, properties, toxicology, applications and perspectives. *Adv Nat Sci Nanosci Nanotechnol*. doi:10.1088/2043-6262-4-3-033001
- Van Phu D, le Quoc A, Duy NN, Lan NT, Du BD, le Luan Q, Hien NQ (2014) Study on antibacterial activity of silver nanoparticles synthesized by gamma irradiation method using different stabilizers. *Nanoscale Res Lett* 9(1):162. doi:10.1186/1556-276x-9-162
- Wang MY, Shen T, Wang M, Zhang D, Chen J (2013) One-pot green synthesis of Ag nanoparticles-decorated reduced graphene oxide for efficient nonenzymatic H₂O₂ biosensor. *Mater Lett* 107:311–314. doi:10.1016/j.matlet.2013.06.031
- Xing Z-C, Chae W-P, Baek J-Y, Choi M-J, Jung Y, Kang I-K (2010) In vitro assessment of antibacterial activity and cytocompatibility of silver-containing PHBV nanofibrous scaffolds for tissue engineering. *Biomacromolecules* 11(5):1248–1253

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